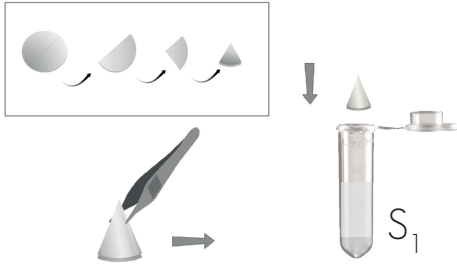




DNA PURE-FLASH

operating mode

EXTRACTION



1. After filtration of the sample, fold the filter in 8 as described above, and place it into the lysis buffer tube (S_1), tip pointing upwards.



2. Incubate at 95°C for 15 minutes to perform thermal lysis. Remove the membrane and discard.



3. Centrifuge the tube 2 minutes at 500 -1000 g, or allow the resin to decant for 5 minutes.

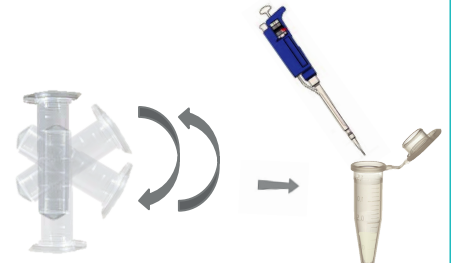
PURIFICATION



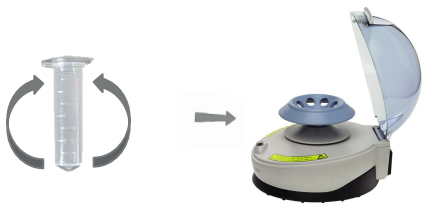
4. Pipet 1 ml of supernatant into a new 2.0 ml tube. Be careful not to sample resin.



5. Add 2 drops of binding buffer (S_2) into the microtube.



6. Manually shake the tube. Then, transfer the entire volume of solution into the WCX resin tube.



7. Stir the tube 20 seconds at 2 000 RPM with a vortex mixer, then centrifuge 30 seconds.



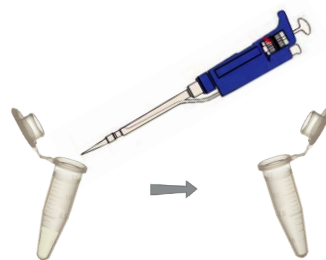
8. Discard the supernatant manually or with a vacuum pump using a tapered tip.



9. Add 100 µl of elution buffer (S_3) in the microtube containing the resin.



10. Mix the microtube using the thermomixer at 1 800 RPM, 75°C ± 5°C for 10 minutes.



11. Centrifuge 1 minute at 2000 g and extract 50 to 100 µl of supernatant. Store at -20°C or analyze directly by qPCR.

ERGONOMY

- Consumables stable at 5°C for more than 6 months
- Analysis performed in under 30 minutes for 5 samples
- Manpower reduced



30 min

EQUIPMENT REQUIRED

- Benchtop micro centrifuge
- Thermomixer
- Micropipettes P200, P1000



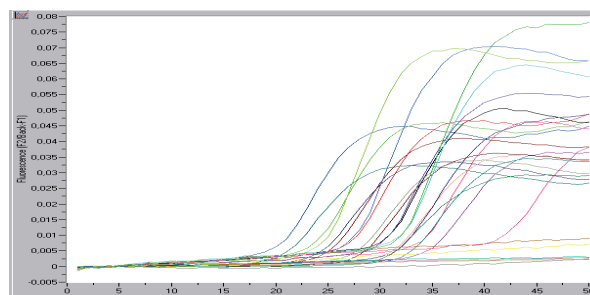
RESULT CALCULATION

$$GU/L = GU_{PCR} \times Z \times 1/V$$

with $Z = 44$

$$GU_{PCR} = GU \text{ quantified by qPCR in } 5\mu l$$

V = Volume of sample filtered in liter



PERFORMANCES

- Recovery higher than 50% on every matrix
- Purification very effective, less than 1% of inhibited sample
- LDminimum (1 liter filtered) = 44 GU/L
- LQminimum (1 liter filtered) = 660 GU/L (with LQ_{pcr} = 20 GU/5 μ l)